Enzyme Computing

第九組 B99901003 楊其昀 B99901071 陳燦帆 B99901123 林鈺翔

DNA Computing

Parallel computing
Faster and smaller
There are problems with scaling
DNAzyme, Enzyme, toehold exchange, Algorithmic self-assembly

Enzyme Computing

- Itamar Willner, 2006
- Insert into body
- Compute metabolic pathway
- Drug delivery
- Monitor the growth of cells

Logic Gates

• Type : AND 、 OR 、 XOR 、 InhibA

\odot Input : GOx \lor GDH \lor AlcDH \lor MP-11

Output : spectral changes

Input Enzymes

• GOx : glucose oxidase

• GDH : glucose dehydrogenase

AlcDH : alcohol dehydrogenase

• MP-11 : microperoxidase-11

Output : Spectral Changes

• ABTS : absorb $\lambda = 420 \ nm$

• NADH : absorb $\lambda = 260 nm \& 340 nm$

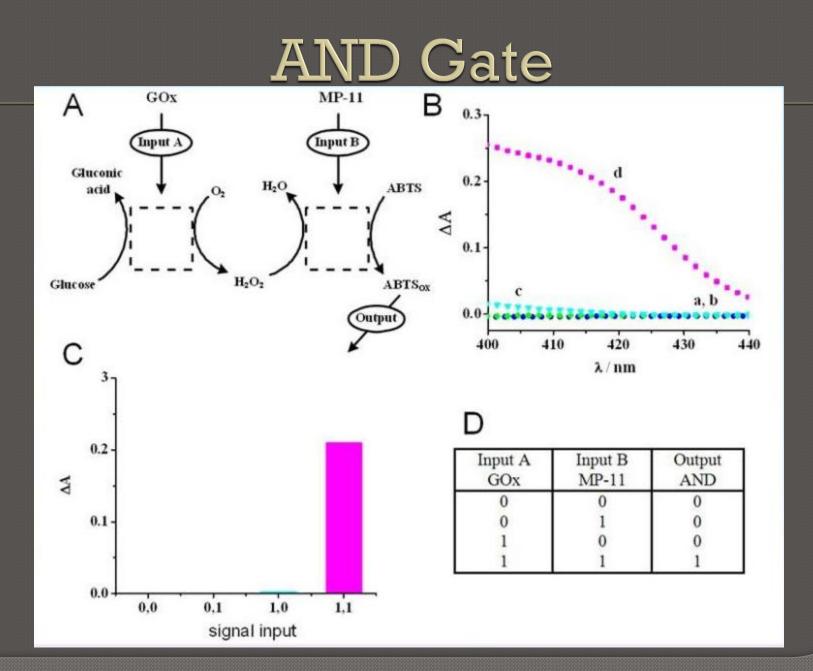
• NAD⁺ : absorb only $\lambda = 260 \ nm$

AND Gate

 Solution containing glucose, oxygen, and ABTS.

• Absorbance $\lambda = 400 \sim 440 \ nm$

 Note that H2O2 does not exists in original composition of the gate

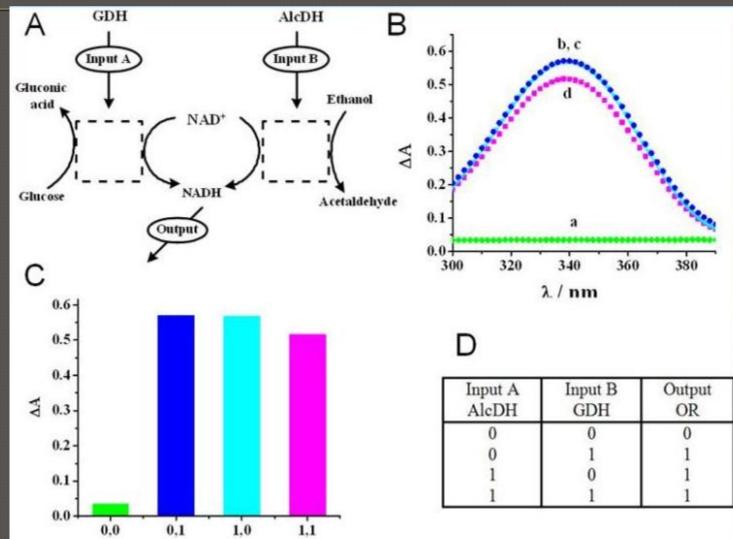


OR Gate

Solution containing glucose, ethanol, and NAD⁺.

• Absorbance $\lambda = 300 \sim 370 \ nm$





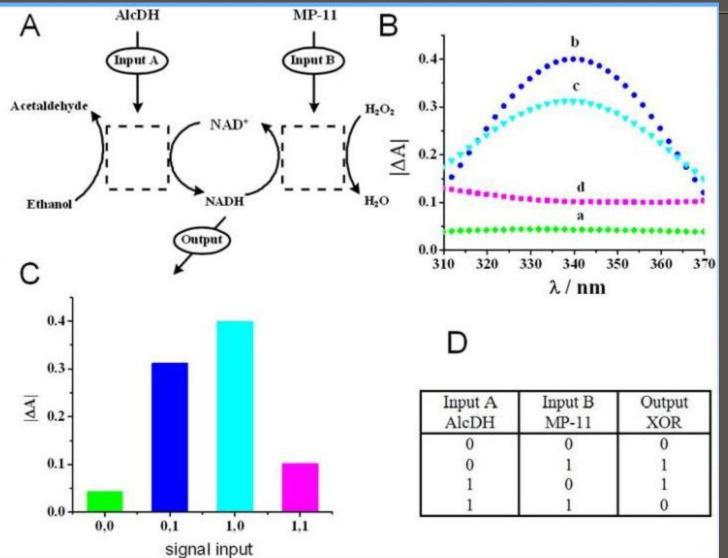
signal input

XOR Gate

• Solution containing H_2O_2 , ethanol, NADH, and NAD^+ .

• Absorbance $\lambda = 300 \sim 370 \ nm$



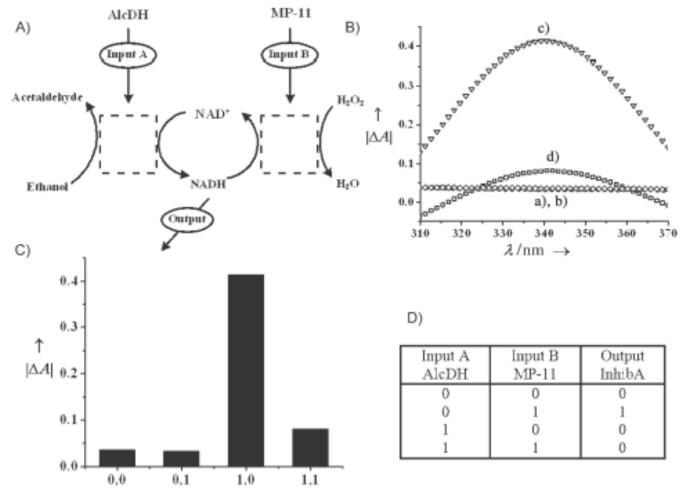


InhibA Gate

• Solution containing H_2O_2 , ethanol, and NADH.

• Absorbance $\lambda = 300 \sim 370 \ nm$

InhibA Gate



signal input

Inverter

• NOR, NAND

• Using different AND & OR

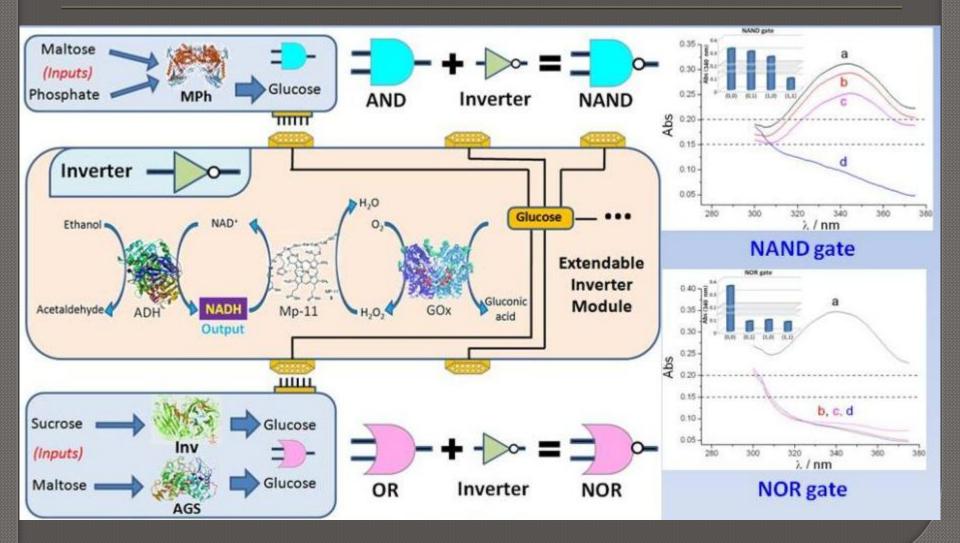
Inverter

 AND : MPh (maltose phosphorylase) pH 7.2, HEPES buffer

 OR : Inv (invertase) AGS (amyloglucosidase) pH7.2, phosephate buffer

Inverter : ADH, GOx, NAD⁺, MP-11 ethanol

Inverter



● Become more complicated → high noise
 level

Concentration

Negative catalyst

• HRP, H_2O_2 , TMB, Ascorbate

Input : H_2O_2 Catalyzed by HRP

Output : TMB and TMB_{ox} \rightarrow blue

Convex function : small input can still cause saturation

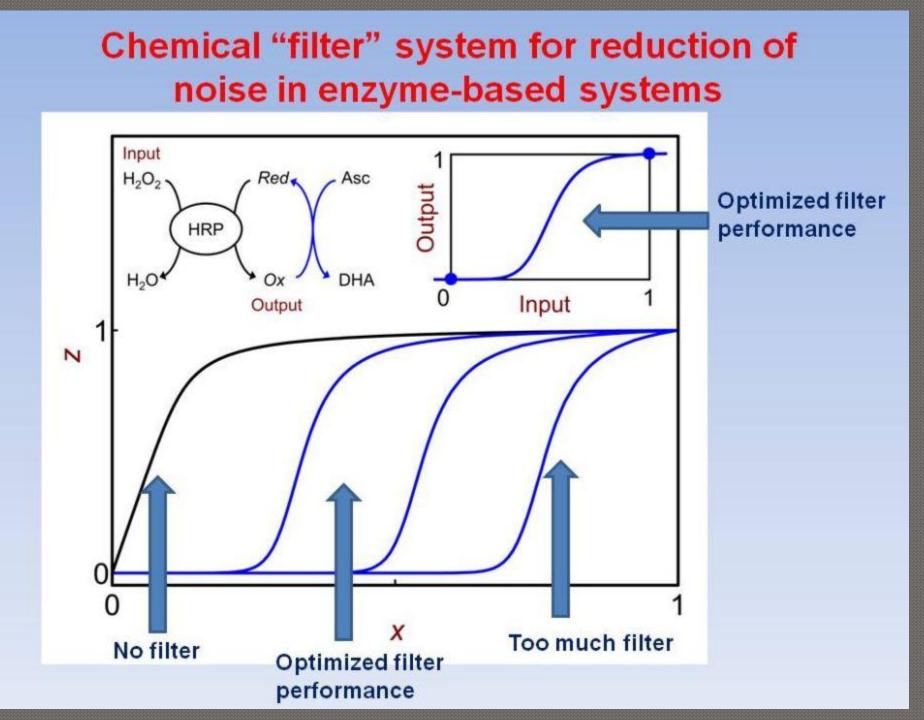
 $\xrightarrow{R} \text{HRP-I} + \text{H}_2\text{O}$ HRP H₂O \xrightarrow{r} HRP-II + TMB^{+·} HRP-I + TMB- $HRP-II + TMB \xrightarrow{r'} HRP + TMB^{+} + H_2O$ $2 \text{TMB}^+ \xrightarrow{r_1} \text{TMB} \cdot \text{TMB}_{ox}$ $TMB \cdot TMB_{ox} \leftarrow TMB + TMB_{ox}$

F: neutralize output
 Ascorbate (ASC)

• Small input : $P \rightarrow 0$ Large input : $P \rightarrow 1$

 $E + I \xrightarrow{R} C$ $C + S \xrightarrow{r} E + P$ $P + F \xrightarrow{\rho} S +$

 $HRP + H_2O_2 \xrightarrow{R} HRP-I + H_2O$ $HRP-I + TMB \xrightarrow{r} HRP-II + TMB^{+}$ HRP-II + TMB $\xrightarrow{r^{*}}$ HRP + TMB⁺⁻ + H₂O $2 \text{TMB}^+ \xrightarrow{r_i} \text{TMB} \cdot \text{TMB}_{ox}$ $TMB \cdot TMB_{ox} \xrightarrow{\prime_2} TMB + TMB_{ox}$ r_{-2} $TMB \cdot TMB_{ox} + Asc \xrightarrow{\rho} 2TMB + \dots$



• Buffer

●Ethyl butyrate(丁酸乙酯) → butyric acid(丁酸)

• Measured by the drop in the pH value.

●Esterase(酯酶)當催化劑

• Small input \rightarrow HEPES(buffer) consumes most of the H⁺

• Large input \rightarrow produced H⁺ ions overwhelm the buffer

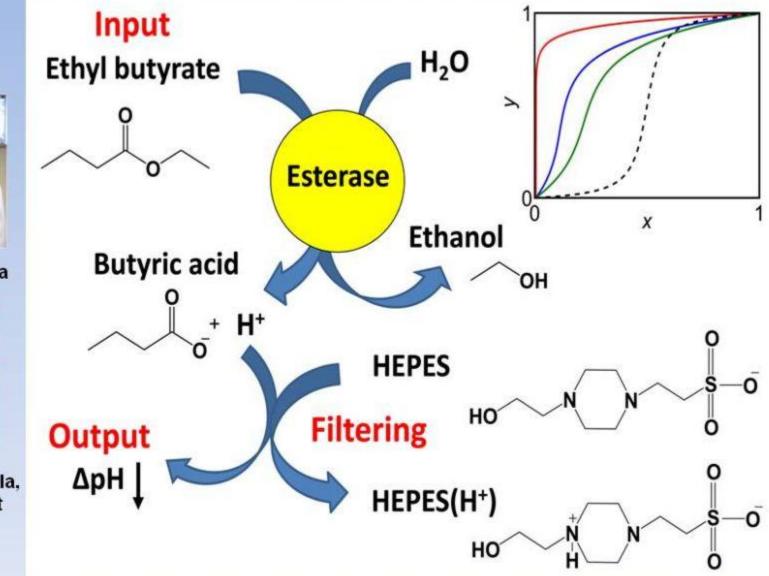
Buffered Biocatalytic Signal Transduction



Dr. Marcos Pita



Mary A. Arugula, PhD student



 Use ADH as the biocatalyst to produce NADH with the presence of NAD⁺ and ethanol.

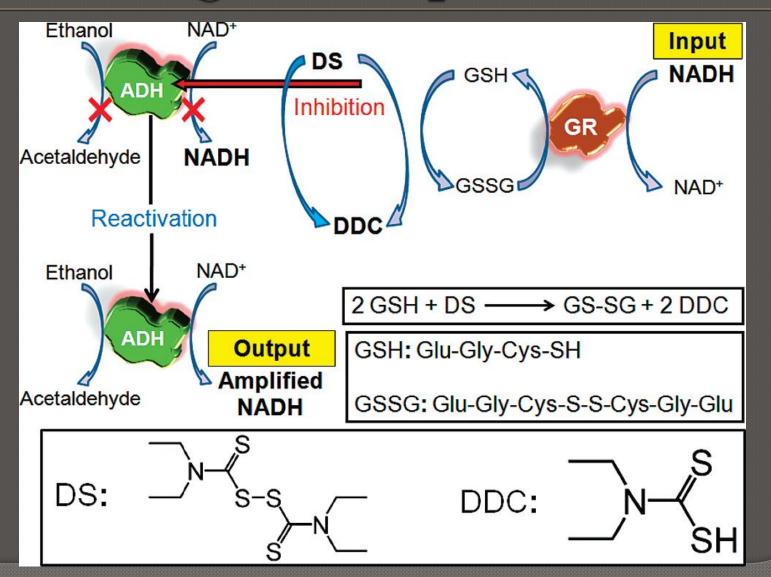
The process must be activated by NADH signal.

• Use DS to inhibit ADH.

Need a system to remove the inhibitor that initialized by NADH.

• GSH can reduce DS and oxidized to GSSG .

With presence of NADH and GR, GSSG reduced to GSH.



• Concentration of DS must be optimized

• Lower: No substantial inhibition of ADH

Higher: Not allow the enzyme reactivation

