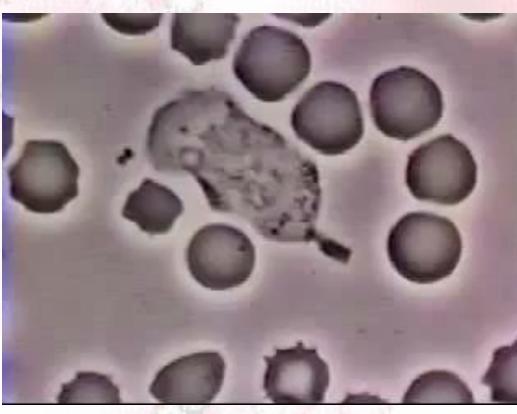
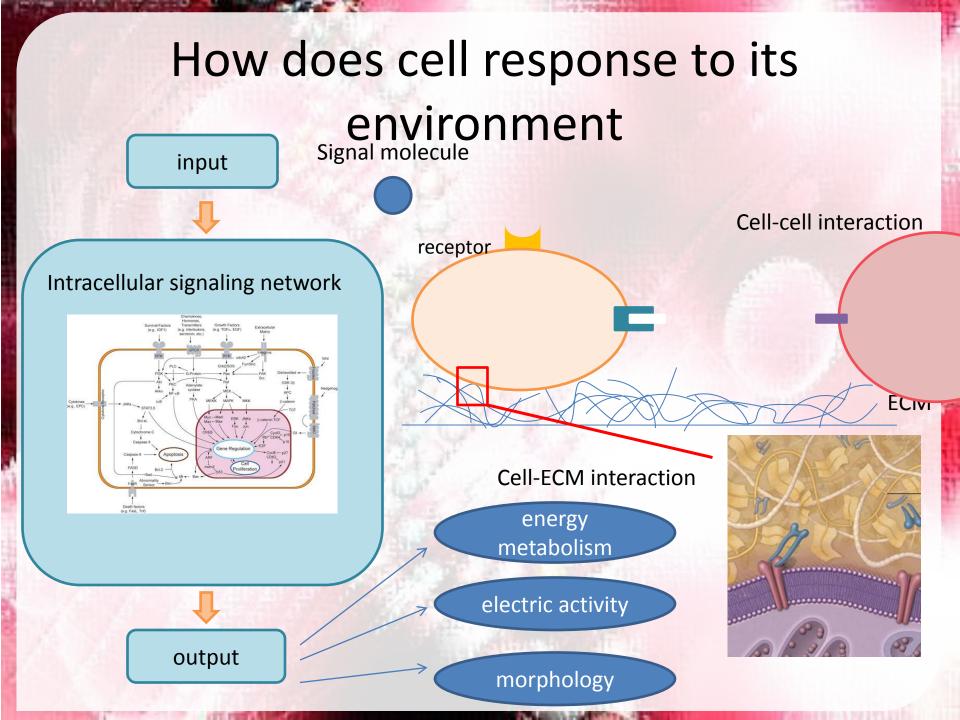


Before we start....



Think about...

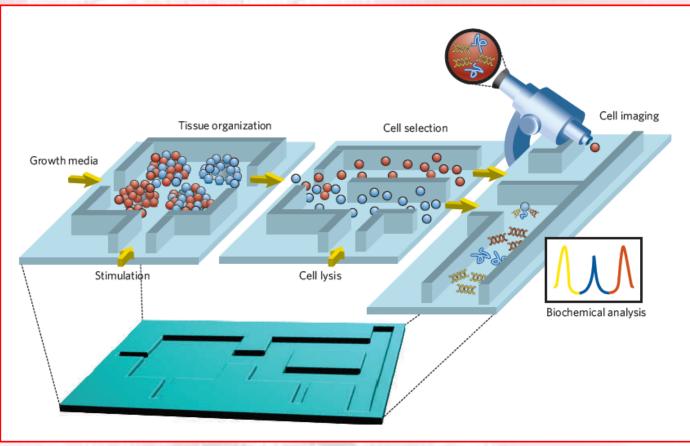
- 白血球細胞沒有大腦思考,如何辨認敵我?
- 如何追蹤細菌?
- 沒有腳如何快速移動?



Cell on chip

- Provides spatial and temporal control of cell growth and stimuli :
 - surfaces that mimic complex biochemistries and geometries of the extracellular matrix
 micro-fluidic channels that regulate transport of fluids and soluble factors

Further integration with bio-analytic microsystems
 1. multifunctional platforms for basic biological insights into cells and tissues



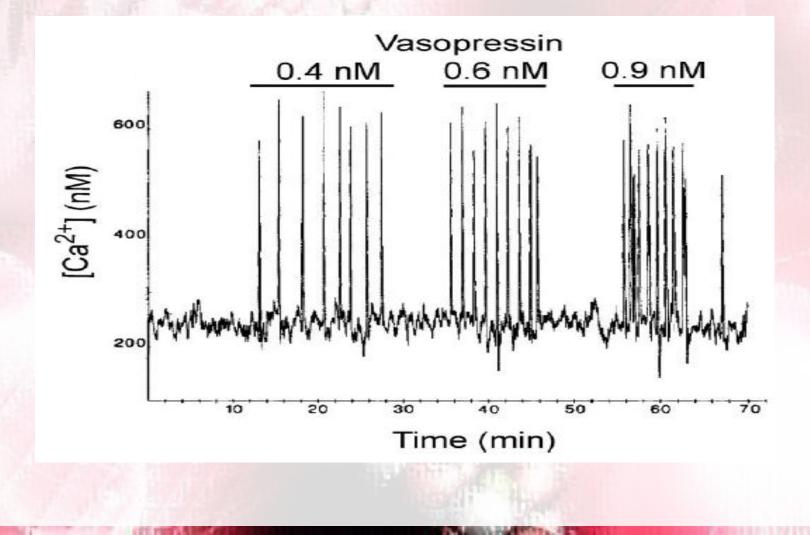
- 2. cell-based sensors
- -- sensing the cell response

single cell analysis

- Most cell-based biological assays yield data averaged across large groups of cells
- Individual cells, even those identical in appearance, differ in numerous characteristics.
- Traditional biochemical assays which analyze cells in bulk often overlook the rich information available when single cells are studied.

example

Cell response after vasopressin stimuli



Applications

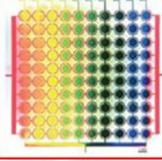
- 1. biological cell-level research
- 2. basic biomedical and pharmaceutical research
- robust and portable point-of-care devices in clinical setting

On-chip analysis of single cells in micro-engineered environments

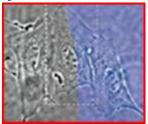
- Control of the fluidic microenvironment
- Gradient control
 控制某種可溶性分子在整個細胞大環
 境的濃度梯度分布
- Local control

控制某種可溶性分子在**單一細胞之周圍環** 境之濃度

Sub-cellular control
 控制某種可溶性分子在單一細胞之不同部分
 的濃度分布







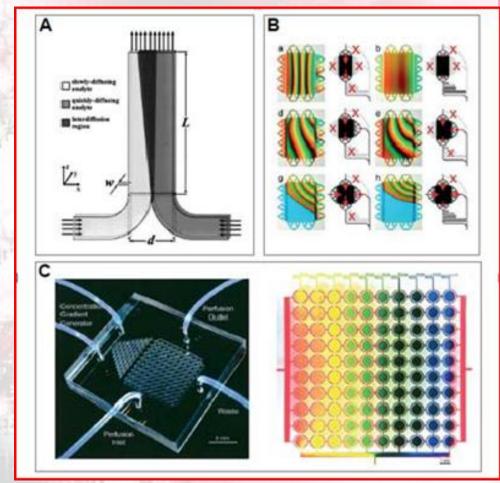
Control of the fluidic microenvironment

Gradients Traditional Methods

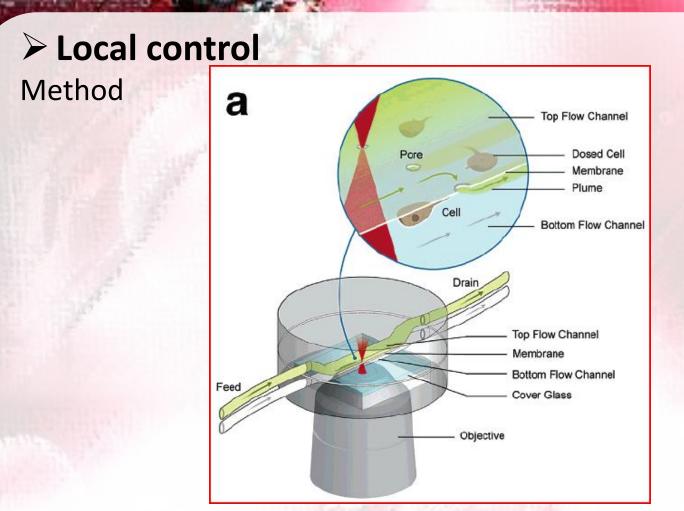


- Poor reproducibility and spatiotemporal control
- Cannot generate complex and stable gradients

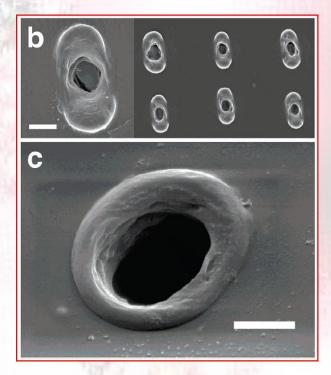
Utilizing laminar flow



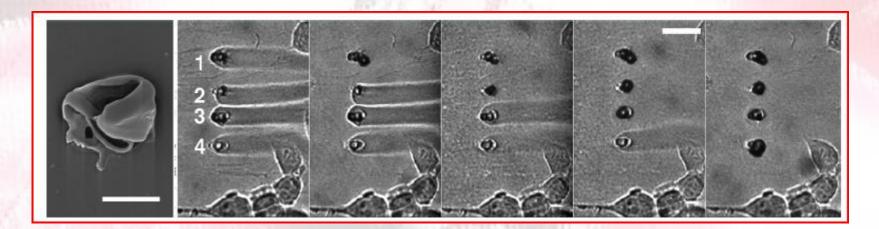
Micro-fluidic gradient generator



- A ultrathin polymer membrane that separates two stacked laminar-flow chambers
- A train of focused femtosecond laser pulses
- Create pores in the membrane producing laminar flow effector stream that enters the lower pressure cell culture chamber.



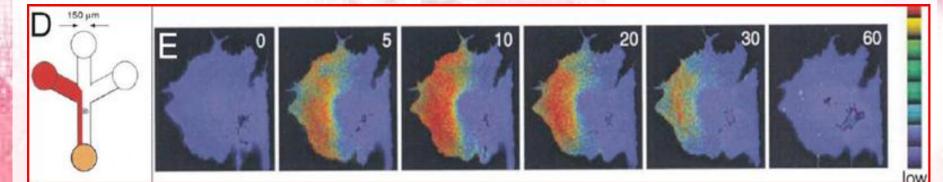
Closure of pores using protein photocross-linking.



Subcellular control - laminar flow

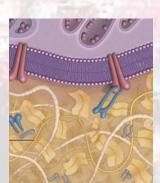
Method

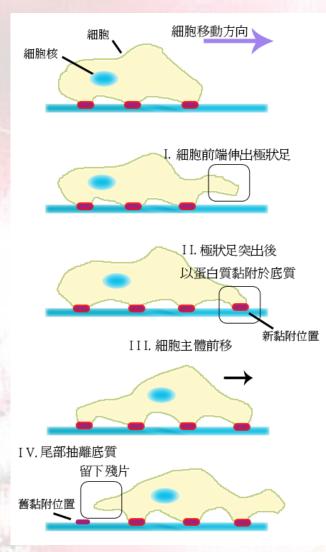
- Utilizing two merging fluid streams
- Expose selective regions of a cell to the soluble hormone epidermal growth factor (critical to cell growth and survival)
 Result
- Normal cells :
 - receptor signaling remained localized
- Cells overexpressing the hormone's receptor : the signal spread throughout the cell ex: a number of tumors



Micro-engineered, three-dimensional substrates

- Cell motility
- Change the arrangement of cytoskeleton with the help of myosin
 - morphology change
- 2. Interaction with ECM
 - movement





Micro-engineered, three-dimensional substrates

Method I : Photolithography

- Using UV light to expose a mask containing the desired patterns onto photosensitive resists.
- Transfer biomolecules of interest by etching or lift-off onto the resist patterns.

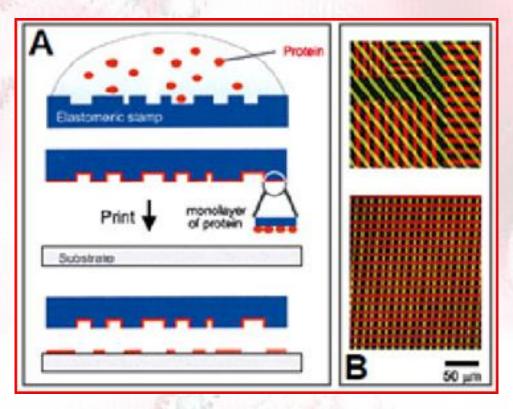
➢ Pros:

pattern quality and patterns with high resolution
Cons:

1. requires cleanroom facilities

2. have to be modified for each new substrate

Method II : Microcontact printing (μCP)



- Microstructured stamp (fabricated in PDMS by molding).
- Coating stamp with molecule of interest by dipping in solution.
- Put the stamp in contact with the substrate to allow transfer.
- Nonprinted adjacent surface can be made passive with another molecule to prevent cell spreading beyond the printed areas.

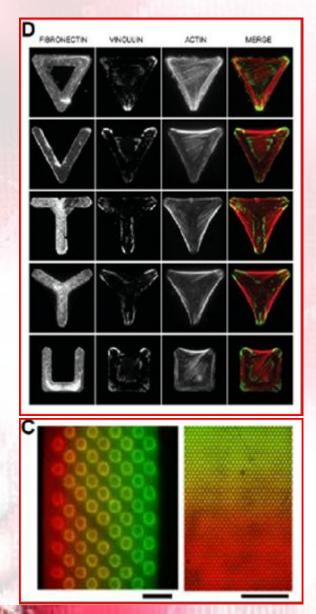
Microcontact printing (µCP)

> Pros :

- Easy and convenient
- Can be used ~100 times over a period of several months without noticeable degradation of the quality of the printing
- Can achieve resolution below 500 nm

Cons:

- Typically is limited to patterns containing only one or two types of molecules.
- > Applications :
- axon guidance
- cell culture on defined geometry



Applications

Tremendous utility in almost all areas of single cell biology ➤ Examples:

- stem cell differentiation
- neuronal regeneration
- cancer metastasis

Cell-based sensor

Ways 1. In real time and non-invasively.
 2. Growing cells directly on the surface sensor chips
 3. Materials which are non-toxic and accepted by most cells as substrate

1. Potentiometric (V)

Metrics

2. Amperometric (I)

3. Impedimetric (R)

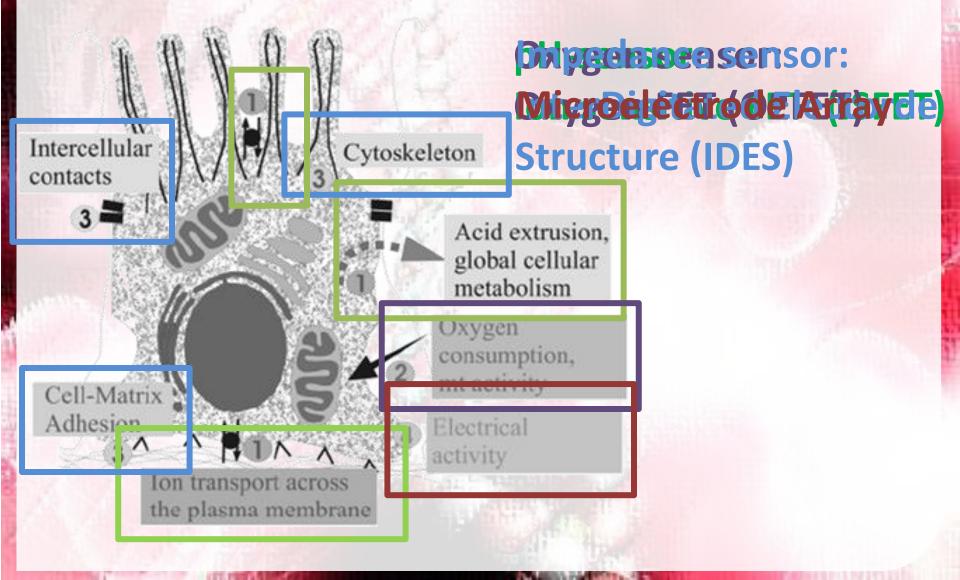
Usages 1. Pharmaceutical drug discovery
 2. Individualized clinical testing
 3. Whole-cell-based biosensors.

Traditional ways to analysis of living cells - Optical screening with dyes

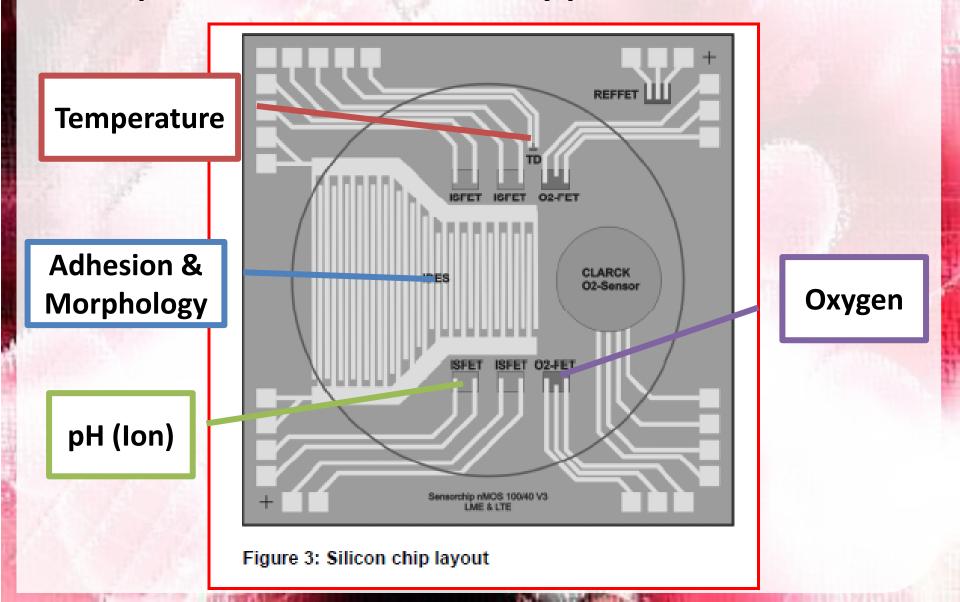
- Toxic or phototoxic properties
- Less suitable for long term monitoring when many dyes applied
- No appropriate dyes for time-resolved studies
 - cell adhesion
 - cell metabolic activity
 - electric activity

(which is the domain of sensor based functional monitoring of cells)

Parameters accessible to microsensor

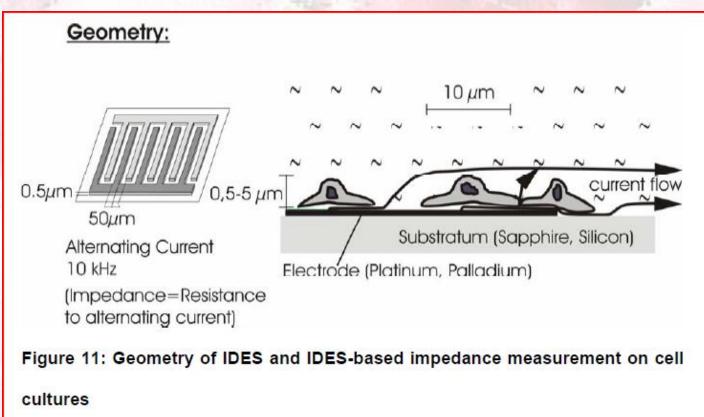


chip with different types of sensors



Sensors Directed Adhesion & Morphology

 changes in cell adhesion or cell morphology result in changes in impedance.



Challenges for Single-cell Analyses on LOC

- The small absolute amount and low concentrations of the cellular species of interest
 - \rightarrow hard to detection.
- Purification or separation strategies prior to analysis
 →loss or dilution
- the mixed hydrophobic—hydrophilic nature of biological macromolecules
 - \rightarrow adsorption to surfaces
- living cells tend to be perturbed by manipulation
 →stringent requirements in performing experiment

Summary

- Cell on chip provides spatial and temporal control of cell growth and stimuli
- As a result, It provides biologists with unprecedented opportunities for cell handling and investigation on a single cell basis
 - Electronic microstructures on sensor chips can analyze cellular responses by recording properties of cell metabolism and morphology

Reference

- "Cells on chips," Jamil El-Ali , Peter K. Sorg and Klavs F. Jensen , *NATURE*, Vol. 442,2006
- "Living cells on chip: bioanalytical applications," Brischwein M, Grothe H, Otto AM, Stepper C, Motrescu E, Weyh T and Wolf B, Mirsky VM (ed) Ul-trathin electrochemical chemo- and biosensors. Springer, Berlin, 159–180.
- "Analysis of single mammalian cells on-chip," C. E. Sims and N. L. Allbritton, Lab Chip, 2007, 7, 423– 440.